Survival and nutritional indexes of *Spodoptera frugiperda* (J.E. Smith, 1797) (Lepidoptera: Noctuidae) maintained in *Bt* maize for five generations

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ABSTRACT: The selection pressure generated by the incorrect use of *Bt* maize might result in *Spodoptera frugiperda* individuals resistant to the toxins synthesized by the plant. In this study was evaluated the nutritional parameters of *Spodoptera frugiperda* submitted to the maize that synthesize the toxins Cry1F, Cry1A105-Cry2Ab2 and Vip3Aa20, during five subsequent generations of selection pressure. The caterpillars were submitted to the treatments: Non-Bt maize (CONV), Cry1F (Bt1), Cry1A105/Cry2Ab (Bt2) and Vip3Aa20 (VIP1, VIP2 and VIP3) and exposed for a period of four days in each cycle, for five successive generations. The larval survival of *S. frugiperda* was evaluated in five generations. The consumed leaf area was quantified in the second generation and the nutritional indexes in the latter ones. The results indicate that caterpillars of *S. frugiperda* from VIPs lineage presented the lowest percentage of survival, with a high Metabolic Cost and a considerable efficiency reduction in the conversion of the digested and ingested food. The lineage maintained in VIP2 showed the lowest leaf consumption (83.13%). However, the action of Vip3Aa20 toxin on *S. frugiperda* resulted in a higher Metabolic Cost in reduction of leaf area consumption. However, from the third generation on there is a survival of insects exposed to *Bt* toxins, suggesting the surviving potential of this species in *Bt* maize subjected to continuous exposure to this technology.

Key words: *Bt* resistance; Cry toxins; fall armyworm; nutritional indexes; Vip3Aa20

Sobrevivência e índices nutricionais de *Spodoptera frugiperda* (J.E. Smith, 1797) (Lepidoptera: Noctuidae) mantida em milho *Bt* por cinco gerações

RESUMO: A pressão de seleção gerada pelo uso incorreto do milho *Bt* pode resultar em indivíduos de *Spodoptera frugiperda* resistentes às toxinas expressadas pela planta. Neste trabalho foi determinado os parâmetros nutricionais de *Spodoptera frugiperda* submetida aos milhos que sintetizam as toxinas Cry1F, Cry1A105-Cry2Ab2 e Vip3Aa20, durante cinco gerações. As lagartas foram submetidas aos tratamentos: Milho não-Bt (CONV), Cry1F (Bt1), Cry1A105/Cry2Ab (Bt2) e Vip3Aa20 (VIP1, VIP2 e VIP3) e expostas por um período de quatro dias em cada ciclo, por cinco gerações sucessivas. Foi avaliada a sobrevivência larval de *S. frugiperda* em cinco gerações. A área foliar consumida foi quantificada na segunda geração e nas posteriores foram determinados os índices nutricionais. As lagartas de *S. frugiperda*, da geração mantida no híbrido VIP2, mostraram menor porcentagem de sobrevivência, com um alto Custo Metabólico e considerável redução na Eficiência na conversão do alimento digerido e Eficiência na conversão do alimento ingerido. A geração mantida no híbrido VIP2 apresentou o menor consumo foliar (83.13%). Contudo a ação da toxina Vip3Aa20 sobre *S. frugiperda* resultou no maior Custo Metabólico e redução no consumo de área foliar. Contudo a partir da terceira geração observa-se uma sobrevivência de insetos expostos as toxinas *Bt*, sugerindo o potencial desta espécie em sobreviver aos milhos *Bt* quando submetidas a exposição contínua a esta tecnologia.

Palavras-chave: resistência *Bt*; toxinas Cry; lagartas-do-cartucho; índices nutricionais; Vip3Aa20
Introduction

A Bt maize contains one or more genes from the entomopathogenic bacterium Bacillus thuringiensis, which synthesizes toxins against the crop main pest, Spodoptera frugiperda (J.E. Smith, 1797) (Lepidoptera: Noctuidae), popularly known as the fall armyworm. Due to the high control rate provided by the Bt technology, farmers grow Bt maize throughout the year, without adopting adequate refuge areas (Storer et al., 2010; Farias et al., 2016).

With the rising use of Bt maize, there is an increase in selection pressure on S. frugiperda, resulting in caterpillars resistant to the toxins synthesized by Bt maize (Storer et al. 2010, Farias et al., 2016, Bernardi et al., 2017). This resistance may be associated with an alteration in the toxin site of action (due to an adaptive response), and/or be linked to the stage of development or even the attacked part of the plant, whereas this site synthesizes a low amount of the toxin (Wang et al., 2004; Santos-Amaya et al., 2017; Yinghua et al., 2017).

Insects resistant to Bt toxins, such as Cry1F toxin, may display alterations in several biological parameters, such as the development time from egg to adult and also reproduction, with an adaptive metabolic cost in response to Cry toxins resistance being noted (Bernardi et al., 2017).

The metabolic cost can be estimated from quantitative nutritional studies, with nutritional indexes being related to the amount of used food, digested, assimilated, metabolized and converted into body material (Scriber & Slansky Jr, 1981). The efficiency in the conversion of digested and ingested food are the main nutritional indexes related to the metabolic cost of S. frugiperda to Bt maize (Yinghua et al., 2017). In Bt cotton that synthesizes the Cry1Ac toxin, the efficiency in the conversion of the digested and ingested food for S. frugiperda was reduced by 9.05 and 8.24%, respectively in relation to the control group (Ramalho et al., 2011).

Reports suggest high evolution risk of the S. frugiperda resistance to pyramidal maize and to the Vip3Aa20 event (Bernardi et al., 2015a; Bernardi et al, 2017). The Bt toxins present in plant leaves can affect the amount of food ingested and converted into body matter, as well as increasing energy expense during metabolism (Ramalho et al., 2011). Thus, the aim of this study was to evaluate the survival and nutritional indexes of Spodoptera frugiperda fed with the maize hybrids that synthesize the toxins Cry1F, Cry1A105-Cry2Ab2 and Vip3Aa20 for five generations.

Materials and Methods

Study site

The study was conducted in the Phytotechny Laboratory, Federal University of Piauí, Professor Cinobelina Elvas Campus, Bom Jesus, PI. The experiment was conducted from November 2012 to October 2013.

Collection of the caterpillars

The insects were collected in conventional and transgenic maize crops (in order to obtain a population with greater genetic variability) at commercial planting areas in the Upper Middle Gurguéia region, Piauí, during the 2012/13 harvest. The collected caterpillars were individually packed in 100 mL plastic containers with a lid.

Creation maintenance

The S. frugiperda caterpillars were sustained with an artificial diet adapted from Kasten Júnior et al. (1978) in a controlled environment (25±2°C; 60±25%RH; 12:12LD). The insects were kept for three generations in order to increase the genetic variability of the population. Thereby, a population with a homogeneous stage of development, with a sufficient number of caterpillars to assemble the bioassays, was obtained in this said generation. Neonates caterpillars (<24h old) were individualized and transferred to 100 mL plastic containers supplied with artificial diet until they reached the pupal stage. The pupae were then transferred to PVC cages (40 cm h x 30 cm Ø), internally covered with sulphite paper sheets, for oviposition. The adults were fed a solution composed by distilled water and honey (10%), and kept at a temperature of 25°C ± 2°C. The egg masses were collected and stored in plastic bags, and kept in controlled environment until they hatched.

Cultivation of maize hybrids

The maize hybrids and their respective toxins used in the experiments were: CONV – non-Bt hybrid (DuPont Pioneer 30F53); Bt1 – Cry1F (DuPont Pioneer 30F53H); Bt2 – Cry1A105-Cry2Ab2 (Syngenta Ag7088); VIP1 – Vip3Aa20 (Syngenta 7205); VIP2 – Vip3Aa20 (Syngenta PENTA); VIP3 – Vip3Aa20 (Syngenta 8A 98). The CONV and Bt1 hybrids are isogenic (same hybrid, but with the introduction of the toxin into Bt1), and the VIPs all have the same insecticidal toxins, in different hybrids.

The selection pressure process was performed using leaves from Bt maize hybrids between 20 and 40 days after the emergence, from plants kept inside a greenhouse in pots with 4 Kg of corrected and fertilized soil, as recommended for the crop (Sousa & Lobato, 2004), without exposure to insecticides. The leaves were collected, sanitized (1% sanitary water solution for 10 minutes), cut and made available for feeding the caterpillars for a period of four days. Confirmation of leaf protein expression was performed by immunodetection tests of the interest proteins using strips from the QuickStix Kit for Vip3A, Cry1F and Cry1A according to instructions of the manufacturer (EnviroLogix Inc. 2013).

Obtaining the lineages of Spodoptera frugiperda

The breeding of the collected insects was held in order to obtain a base population with greater genetic variability. The progenies of these breeds were divided into sub-populations or lineages. The caterpillars remained on artificial diet until the third instar, and later were fed maize leaves, where each hybrid was considered as a treatment and as acorresponding to a lineage. One of the lineages was kept in the absence of selection pressure in conventional maize leaves. The others
were submitted to the transgenic events containing the proteins Cry1F, Cry1A105/Cry2Ab and VIP3AA20. The caterpillars were exposed for four days in each generation, for five generations. The principle of low selection pressure proposed by Roush & McKenzie (1987) was adopted, where the high dose makes the expression of the resistance effectively or functionally recessive, eliminating the heterozygous individuals, with the low being able to cause an effectively dominant resistance.

Surviving insects, after this period, were kept on an artificial diet in a controlled environment as previously described, until they completed the cycle.

**Evaluated parameters**

The nutritional indexes from the different lineages of *S. frugiperda* were determined using the fresh weight of the leaves, caterpillars and feces of the insects, where they fasted for 15 hours prior to that. Afterwards, the caterpillars were weighed and fed with the leaves of the previously weighed hybrids. After four days, another weight of the caterpillars, leaves and feces was done, following the proposed methodology by Waldbauer (1968), modified by Scriber & Slansky Jr (1981). Eight replications (10 caterpillars/replicate) of each lineage were used, kept for four days in maize leaves. The calculated parameters were: T = duration of the feeding period (days); Af = weight of food given to the insect (g); Ar = weight of the leftover food given to the insect (g), after T; F = weight of produced feces (g) during T; b = caterpillars weight gain (g) during T; B = caterpillars mean weight (g) during T; I = weight of the ingested food (g) during T; I - F = assimilated food (g) during T; M = (I - F) - B = metabolized food during the feeding period. For the weights measurement, a precision analytical balance was employed. The calculated consumption indexes were:

- Relative consumption rate (RCR) = I/B*T;
- RGR (Relative growth rate = b/B*T);
- RMR (Relative metabolic rate = M/B*T);
- AD (approximate digestibility = I-F/I*100);
- ECI (Conversion efficiency of ingested food = b/I*100);
- ECD (Conversion efficiency of digested food = b/I-F*100);
- CM (metabolic cost = 100 - ECD).

The leaf area was estimated using the LI – 3.100C Area Meter (LI-COR™ Biosciences) equipment, in the second generation of individuals. Fresh leaves were cut, measured and offered to the caterpillars, and at the end of the four-day experiment time, the leaves were measured again, where the difference between the readings corresponded to the leaf area consumed by the caterpillar.

**Statistical analysis**

The bioassay was conducted in a completely randomized design (DIC) consisting of five treatments and eight replicates each. Data regarding the mortality were transformed to x cotangent, in radians, and subjected to analysis of variance (Proc GLM). When found a significant difference, the results were submitted to the Duncan mean comparison test, and the accumulated mortality index per generation adjusted with eight replicates for each treatment within the five generations, with 80 insects per treatment (10 caterpillars/replicate).

The nutritional indexes were transformed into arch of x cotangent, in radians, and subsequently subjected to the individual analysis of variance (per generation) (Proc GLM), where the variances were compared by Fisher test and, after homogeneity verification of the residual variances, the joint analysis was held. When significant difference was detected for the interaction, the unfolding within and between generations was performed using Duncan mean test.

The Pearson correlation coefficient was determined using nutritional indexes within each generation. In the second generation, the leaf area consumed by the caterpillars was evaluated, where 32 replicates were held for each hybrid. Analyzes were performed by PROC GLM, and the Duncan means comparison test. Statistical analyzes were performed with the SAS program (SAS, 2002).

**Results**

Caterpillars fed with Bt maize hybrids had larval survival reduced over the five generations. The mean survival of *S. frugiperda* within each generation when fed with the hybrids CONV, Bt1, Bt2, VIP1, VIP2, VIP3 showed a statistical difference (Figure 1).

The survival percentage of *S. frugiperda* caterpillars kept without selection pressure and fed with non-Bt maize remained relatively constant over the generations. Among the genetically modified hybrids, survival was highest in Bt hybrids, up to the third generation, when compared to VIP hybrids. From the third generation on, the survival was rising in all Bt hybrids. From the fourth generation of breeding on, there was a survival increase in the Bt2 and VIP1 lineages, while in the others we observed a slight decrease. VIP2 and VIP3 reduced insect survival within the evaluated generations by 34.38% and 28.13%, respectively.

![Figure 1](image_url)

**Figure 1.** Survival of *S. frugiperda* in five generations during selection pressure. (F = 6.31, G.L. = 18, 208, P = 0.01, C.V. 42.97%). CONV – (non-Bt hybrid); Bt1 – (Cry1F); Bt2 – (Cry1A105-Cry2Ab2); VIP1 – (Vip3Aa20); VIP2 – (Vip3Aa20); VIP3 – (Vip3Aa20).
Results regarding nutritional indexes of *S. frugiperda* showed statistical significance (Table 1). Due to problems regarding vigor and germination of the Bt1 hybrid, its nutritional indexes were not estimated. The metabolized food (M) was lower in the third and fifth generations of *S. frugiperda* in CONV and VIP lineages populations, respectively. Caterpillars exposed to the VIP3 hybrid presented a relative consumption rate (RCR) similar to the conventional lineage insects. In the fifth generation, the insects kept in VIP1 hybrid had the lowest relative Consumption Rate (Table 1). Throughout the generations, the relative consumption rate remained similar to that displayed in the third generation by the CONV lineage caterpillars.

Treatments caused an alteration in the relative growth rate (RGR). Within the third generation, only the CONV lineage caterpillars presented growth when compared to the others. In the fifth generation, the CONV and Bt2 lineages caterpillars also maintained the growth. These results suggest a nutritional adaptation by the caterpillar to the Cry1A105-Cry2Ab2 toxin over the generations. Regarding the relative metabolic rate (RMR), the caterpillars kept in the CONV and VIP1 hybrids had the lowest metabolic rates in the third and fifth generations, respectively. In the fifth generation, the caterpillars from VIP2 and VIP3 lineages presented a high relative consumption with a decrease in the relative growth rate and reduced efficiency in the conversion of the ingested and digested food, resulting in a high metabolic cost.

According to the Pearson correlation coefficients (Table 2), we can note that in the third generation, there is negative correlation between the approximate digestibility and the growth rate and relative consumption indexes, with the RCR showing a positive correlation with RGR and ECI. Where the conversion efficiency of the ingested food present a positive correlation with RGR and ECD. For the fifth generation, we observed that the lower the RGR, ECI and ECD values, the higher the metabolic cost is (Table 2). We can also observe that the CONV variety had the largest consumption of leaf area in the second generation (Figure 2). Contrasting to this, the VIP hybrids had a greater reduction in leaf consumption, with emphasis the VIP1 and VIP2 hybrids, with about 80% less consumption than the control group.

**Significant at 1% probability by F test. Means followed by the same uppercase letter in the column and lowercase in the line do not differ significantly from each other by the Duncan test at 5% probability. RCR, RGR, RMR are in g/g/day; and AD, ECI, ECD and CM in %. SE: Mean standard error.**

<table>
<thead>
<tr>
<th>Treatments</th>
<th>3rd Generation</th>
<th>5th Generation</th>
<th>3rd Generation</th>
<th>5th Generation</th>
</tr>
</thead>
<tbody>
<tr>
<td>Hybrid</td>
<td>Metabolized food (M)</td>
<td>Approximate digestibility (AD)</td>
<td>Metabolized food (M)</td>
<td>Approximate digestibility (AD)</td>
</tr>
<tr>
<td>CONV</td>
<td>-0.006 ± 0.001 Aa</td>
<td>0.141 ± 0.02 Ab</td>
<td>49.1 ± 4.18 Ba</td>
<td>81.73 ± 3.66 Ab</td>
</tr>
<tr>
<td>Bt2</td>
<td>0.032 ± 0.01 Ab</td>
<td>0.086 ± 0.007 Aa</td>
<td>76.7 ± 6.41 Aa</td>
<td>82.04 ± 3.08 Aa</td>
</tr>
<tr>
<td>VIP1</td>
<td>0.053 ± 0.02 Aa</td>
<td>0.054 ± 0.008 Ba</td>
<td>77.7 ± 5.64 Aa</td>
<td>89.46 ± 5.1 Aa</td>
</tr>
<tr>
<td>VIP2</td>
<td>0.080 ± 0.03 Aa</td>
<td>0.105 ± 0.004 Aa</td>
<td>86.1 ± 4.92 Aa</td>
<td>99.88 ± 0.12 Aa</td>
</tr>
<tr>
<td>VIP3</td>
<td>0.047 ± 0.004 Aa</td>
<td>0.117 ± 0.014 Aa</td>
<td>87.6 ± 2.95 Aa</td>
<td>98.81 ± 0.27 Aa</td>
</tr>
<tr>
<td>F</td>
<td>5**</td>
<td>3.39</td>
<td>6**</td>
<td>27.34</td>
</tr>
<tr>
<td>C.V.%</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Relative consumption rate (RCR)</td>
<td>Conversion efficiency of ingested food (ECI)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>CONV</td>
<td>3.067 ± 0.12 Aa</td>
<td>3.16 ± 0.63 Ba</td>
<td>5.234 ± 0.74 Aa</td>
<td>8.403 ± 1.19 Aa</td>
</tr>
<tr>
<td>Bt2</td>
<td>1.305 ± 0.16 Ba</td>
<td>4.54 ± 1.61 Bb</td>
<td>-5.703 ± 3.28 Ba</td>
<td>5.795 ± 1.63 Ab</td>
</tr>
<tr>
<td>VIP1</td>
<td>1.541 ± 0.29 Ba</td>
<td>1.78 ± 0.52Cb</td>
<td>-11.247 ± 2.50 Ba</td>
<td>-4.884 ± 0.85 Ba</td>
</tr>
<tr>
<td>VIP2</td>
<td>1.385 ± 0.39 Ba</td>
<td>11.32 ± 1.95 Aa</td>
<td>-14.114 ± 2.38 Ba</td>
<td>-1.151 ± 0.31 Ba</td>
</tr>
<tr>
<td>VIP3</td>
<td>2.389 ± 0.38 Aa</td>
<td>9.43 ± 1.24 Aa</td>
<td>-2.342 ± 0.51 Ba</td>
<td>-1.346 ± 0.41 Ba</td>
</tr>
<tr>
<td>F</td>
<td>11.90**</td>
<td>4.39**</td>
<td></td>
<td></td>
</tr>
<tr>
<td>C.V.%</td>
<td>39.80</td>
<td>33.42</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Relative growth rate (RGR)</td>
<td>Conversion efficiency of digested food (ECD)</td>
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<td></td>
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</tr>
<tr>
<td>CONV</td>
<td>0.160 ± 0.02 Aa</td>
<td>0.195 ± 0.02 Aa</td>
<td>12.204 ± 2.32 Aa</td>
<td>11.021 ± 1.86 Aa</td>
</tr>
<tr>
<td>Bt2</td>
<td>-0.034 ± 0.04 Ba</td>
<td>0.157 ± 0.05 Ab</td>
<td>-4.026 ± 5.26 Ba</td>
<td>7.591 ± 2.03 Ab</td>
</tr>
<tr>
<td>VIP1</td>
<td>-0.123 ± 0.02 Ba</td>
<td>-0.067 ± 0.01 Ba</td>
<td>-15.703 ± 3.52 Ca</td>
<td>-6.241 ± 1.67 Ba</td>
</tr>
<tr>
<td>VIP2</td>
<td>-0.140 ± 0.01 Ba</td>
<td>-0.095 ± 0.01 Ba</td>
<td>-16.401 ± 2.41 Aa</td>
<td>-1.153 ± 0.31 Ba</td>
</tr>
<tr>
<td>VIP3</td>
<td>-0.053 ± 0.01 BCa</td>
<td>-0.107 ± 0.03 Ba</td>
<td>-2.541 ± 0.54 Ba</td>
<td>-1.361 ± 0.42 Ba</td>
</tr>
<tr>
<td>F</td>
<td>46.2**</td>
<td>4.5**</td>
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<td></td>
</tr>
<tr>
<td>C.V.%</td>
<td>5.64</td>
<td>34.19</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Relative metabolic rate (RMR)</td>
<td>Metabolic cost (CM)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>CONV</td>
<td>-0.069 ± 0.26 Ca</td>
<td>2.266 ± 0.64 Ab</td>
<td>87.796 ± 2.32 Ca</td>
<td>88.979 ± 1.86 Ba</td>
</tr>
<tr>
<td>Bt2</td>
<td>0.562 ± 0.16 Ba</td>
<td>3.593 ± 1.68 Ab</td>
<td>104.026 ± 5.26 Ba</td>
<td>92.409 ± 2.04 Ba</td>
</tr>
<tr>
<td>VIP1</td>
<td>1.044 ± 0.32 AbA</td>
<td>1.503 ± 0.56 Ba</td>
<td>115.703 ± 3.52 Aa</td>
<td>106.241 ± 1.67 Aa</td>
</tr>
<tr>
<td>VIP2</td>
<td>1.146 ± 0.42 Aa</td>
<td>11.30 ± 1.95 Ab</td>
<td>116.481 ± 2.42 Aa</td>
<td>101.153 ± 0.31 AbB</td>
</tr>
<tr>
<td>VIP3</td>
<td>1.806 ± 0.25 Aa</td>
<td>9.228 ± 1.24 Ab</td>
<td>102.541 ± 0.54 AbA</td>
<td>101.361 ± 0.42 AbA</td>
</tr>
<tr>
<td>F</td>
<td>4.62**</td>
<td>2**</td>
<td></td>
<td></td>
</tr>
<tr>
<td>C.V.%</td>
<td>51.51</td>
<td>8.67</td>
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</tr>
</tbody>
</table>
of populations resistant to the Cry1F toxin are already documented in the literature (Storer et al., 2010). Survival of at least 35% of *S. frugiperda* in the Bt Cry1F maize (Bernardi et al., 2015b; Farias et al., 2016; Yang et al., 2016).

There was a reduction in the control efficiency in pyramided (Cry1A105-Cry2Ab2) and non-pyramided (Cry1F) events. This finding contradicts the premise that transgenic crops that have two or more toxins would be more effective in delaying the resistance evolution when compared to crops with only one toxin (Tabashnik & Gould, 2012), due to the occurrence of cross-resistance (Bernardi et al., 2015b). Resistant insects possess the ability to eliminate Bt toxins from their intestine, in addition to altering the way Cry toxins act, invalidating the toxic action (Pérez-Hedo et al., 2012).

The significant increase in the survival of caterpillars exposed to hybrids with Vip3a20 technology within three generations corroborates with the obtained results by Miraldo et al. (2016), where the exposure of *S. frugiperda* to Vip3Aa20 maize resulted in the survival of around 20% of caterpillars.

Bernardi et al. (2015a) cite that, due to technology misuse, the fall armyworm may develop resistance to Vip3Aa20 toxins. However, the survival percentages of caterpillars exposed to the used transgenic maize hybrids may be tied to the original crops, where the population used in this study was in the third generation and the insects can maintain resistance in environments without selection pressure for up to seven generations (Santos-Amaya et al., 2017).

The caterpillars exposed to Bt hybrids display altered biological characteristics when compared to insects kept in non-Bt maize hybrids (Bernardi et al., 2017). The nutritional indexes of lineages kept on transgenic maize leaves indicate metabolic cost (CM) and a marked reduction in leaf consumption.

The recorded survival percentage since the first generation indicates that the fall armyworm already had some response variability to the Cry1F and Cry1A105-Cry2Ab2 toxins expressed in the studied maize hybrids, and consequently no change in the insect behavior was observed. Reports of populations resistant to the Cry1F toxin are already documented in the literature (Storer et al., 2010). Survival of at least 35% of *S. frugiperda* in the Bt Cry1F maize (Bernardi et al., 2015b; Farias et al., 2016; Yang et al., 2016).

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**Discussion**

The selection pressure on *S. frugiperda* brought by the continuous use of Bt maize, without having a refuge crop, can condition the selection of resistant insects, resulting in a reduced efficiency of the technology employed (Storer et al., 2010; Farias et al., 2016). Evaluation by successive generations, in this study, resulted in a greater survival of individuals after five generations. High survival was also verified after ingestion of the Cry toxins, although having a metabolic cost (CM) and a marked reduction in leaf consumption.

The recorded survival percentage since the first generation indicates that the fall armyworm already had some response variability to the Cry1F and Cry1A105-Cry2Ab2 toxins expressed in the studied maize hybrids, and consequently no change in the insect behavior was observed. Reports of populations resistant to the Cry1F toxin are already documented in the literature (Storer et al., 2010). Survival of at least 35% of *S. frugiperda* in the Bt Cry1F maize (Bernardi et al., 2015b; Farias et al., 2016; Yang et al., 2016).

There was a reduction in the control efficiency in pyramided (Cry1A105-Cry2Ab2) and non-pyramided (Cry1F) events. This finding contradicts the premise that transgenic crops that have two or more toxins would be more effective in delaying the resistance evolution when compared to crops with only one toxin (Tabashnik & Gould, 2012), due to the occurrence of cross-resistance (Bernardi et al., 2015b). Resistant insects possess the ability to eliminate Bt toxins from their intestine, in addition to altering the way Cry toxins act, invalidating the toxic action (Pérez-Hedo et al., 2012).

The significant increase in the survival of caterpillars exposed to hybrids with Vip3a20 technology within three generations corroborates with the obtained results by Miraldo et al. (2016), where the exposure of *S. frugiperda* to Vip3Aa20 maize resulted in the survival of around 20% of caterpillars. Bernardi et al. (2015a) cite that, due to technology misuse, the fall armyworm may develop resistance to Vip3Aa20 toxins. However, the survival percentages of caterpillars exposed to the used transgenic maize hybrids may be tied to the original crops, where the population used in this study was in the third generation and the insects can maintain resistance in environments without selection pressure for up to seven generations (Santos-Amaya et al., 2017).

The caterpillars exposed to Bt hybrids display altered biological characteristics when compared to insects kept in non-Bt maize hybrids (Bernardi et al., 2017). The nutritional indexes of lineages kept on transgenic maize leaves indicate metabolic cost (CM) and a marked reduction in leaf consumption.

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to the development in question (Horikoshi et al., 2016). We verified in this study that the conversion of ingested, digested (ECI, ECD) and relative growth (RGR) showed a strong negative correlation with the metabolic cost (CM). Usually, organisms exposed to Bt maize shows high metabolic cost in direct response to the toxins action (Ramalho et al., 2011; Cataño et al., 2014). The estimated nutritional indexes were affected by the evaluated toxins action, as well as the obtained results with the exposure of S. frugiperda to Bt cotton (Ramalho et al., 2011; Cataño et al., 2014).

The metabolic cost of caterpillars grown on VIP maize (Vip3Aa20 event) was similar to S. litura fed with Bt maize (Cry1Ab). In this case, there was high consumption without weight gain, where we noted high levels of digested feed (AD) and relative consumption rate (RCR) and low feed conversion efficiency (ECI), digested feed conversion (ECD) and relative growth rate (RGR), indicating an energy shift from biomass production to toxin detoxification (Yinghua et al., 2017).

The changes in metabolized feed (M), relative growth rate (RGR), relative metabolic rate (RMI) and relative consumption rate (RCR) are the result of a deficiency in assimilation and subsequent conversion of nutrients into body material and the inactivation of Bt toxins (Ramalho et al., 2011; Yinghua et al., 2017). In virtue of the presence of toxic substances without interference in palatability, caterpillar intoxication occurs due to the action of Cry toxins, affecting the usage of ingested food (Scriber & Slansky, 1981; Jiang et al., 2013). Thus, the ability of an organism to convert nutrients and protein will influence positively its growth (Sogbesan & Ugwumba, 2008).

The lower values of Approximate Digestibility (AD) obtained in S. frugiperda not exposed to Bts toxins may be related to fast food digestibility in the intestine of the insect (Ramalho et al., 2011). The rapid transition of food through the digestive tract of the insect reduces the interaction of proteolytic enzymes in the food substrate, causing a decrease in the active toxins in the intestinal lumen (Dinglasan et al., 2009). On the other hand, caterpillars fed with Bt maize, mainly those that synthesize the Vip3Aa20 toxin, presented high values of approximate digestibility, indicating that the toxin inactivation affects the food digestibility, since the Cry toxins action in the intestine of the insects alter the activity of the digestive enzymes (Yinghua et al., 2017).

Changes in the conversion efficiency of ingested and digested food may be related to the presence of Vip3Aa20, as the action of Bt toxins reduces the caterpillars ability in assimilating the nutrients (Cataño et al., 2014), or the resulting energy from assimilation of food is being used in the regeneration of midgut epithelium, damaged by the Cry toxins action (Lüthy & Wolfersberger, 2000). Caterpillars grown with VIP maize (Vip3Aa20 event) presented the highest values of metabolic cost, probably due to an inefficient conversion of biomass with low efficiency values in the conversion of ingested and digested food (Cataño et al., 2014).

When comparing the generations, in the third generation the CONV lineage caterpillars and in the fifth generation the Bt2 lineage (Cry1A105-Cry2Ab2 event) presented an increase in the rates of conversion efficiency of the ingested and digested food. Thus, nutritional indexes (especially ECD and ECI) can be considered important within the parameters related to the survival of insects exposed to Bt maize.

CONV and VIP hybrids presented the highest and lowest leaf area consumed, respectively. The presence of Vip3Aa20 toxin in the digestive tract of the insect may have led to midgut partial paralysis, resulting in a decrease in leaf consumption and a considerable metabolic cost (Prütz & Dettner, 2004; Storer et al., 2010; Farias et al., 2016).

The Cry1F-Cry1A105-Cry2Ab2 pyramidal event caused a reduction of 55.14% in the leaf consumption of Spodoptera eridania (Cramer) (Lepidoptera: Noctuidae), in relation to the control group (Bortolotto et al., 2015), results higher than those found in this study, where maize with two proteins (Cry1A105-Cry2Ab2) had a reduction of 32.93% in the consumed leaf area. A low response in the conversion of ingested and deferred food by the insects may result in a greater consumption of leaf area; however, further studies are necessary in order to consolidate this premise. The results contrast with the literature premises, that pyramidal events present a higher percentage of control, resistance prevention and lower leaf consumption (Bortolotto et al., 2015). Costa et al. (2006), observed a 28.92 cm².day⁻¹ consumption of S. frugiperda in leaves of conventional maize, compatible with the result found in this study.

According to the obtained results, it can be established that Bt maize hybrids containing one or more toxins affect the nutritional indexes and the consumed leaf area. In addition to providing subsidy for new studies regarding the activity of the detoxifying enzymes present in the intestines of insects for a better understanding of the Bt toxins toxic effects, studies of cross-resistance to other toxins, which will help in the selection of these for pyramiding in maize plants and/or for the rotation of maize hybrids expressing different toxins. As well as the genetic, biochemical and molecular characterization of lineage resistance, this may aid in the refinement of recommendations for the management of S. frugiperda resistance to transgenic maize with Cry1F, Cry1A105-Cry2Ab2 and Vip3Aa20.

**Conclusion**

*Spodoptera frugiperda* grown with Vip3Aa20 maize presents a survival percentage increase from the second generation onwards.

The relative growth rate, the conversion efficiency of the ingested and digested food present a negative correlation with the metabolic cost in populations of *S. frugiperda* maintained in Bt maize hybrids.

Vip3Aa20 maize carries the highest metabolic cost with reduction in leaf consumption of *Spodoptera frugiperda*.

Bt maize hybrids with the Cry1A105-Cry2Ab2 and Vip3Aa20 toxins present low survival percentages with changes in nutritional indexes and can be employed as an alternative in the *Spodoptera frugiperda* management.
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Literature Cited


